

The James



THE OHIO STATE UNIVERSITY
COMPREHENSIVE CANCER CENTER

Lynn O'Donnell, PhD

Director, Cell Therapy Laboratory
Associate Professor, Division of Hematology

lynn.odonnell@osumc.edu

20th ISCT Annual Meeting, Paris, France

*Quality & Operations Track 1: Cord Blood
Infusions: Current Gold Standards in
Preparing and Testing of Cord Blood
Products for Transplantation*

April 24, 2014



Creating a cancer-free world. *One person, one discovery at a time.*

Impact of Cord Thaw Method on Processing and Clinical Outcome:

Why Choosing the Right Thaw Method Could Save a Patient's Life

The James



THE OHIO STATE UNIVERSITY
COMPREHENSIVE CANCER CENTER

OSU-James BMT Program – Cord Blood Transplants

- Approximately 250 transplant / year
- Late entry into cords – first patient **March 2008**
- Only 4 patients transplanted off-study first year, then opened two clinical trials and accrual picked up
 - BMT CTN 0604 / CIBMTR 05-DCB – adult, nonmyeloablative, double cord transplant
 - BMT CTN 0501 – pediatric, myeloablative, randomized single vs. double cord transplant
- RBC-replete (containing) CBUs were initially preferred due to higher TNC dose
- **April 2009:** Our 2nd patient on 05-DCB experienced SAE and 9 days later the 3rd did as well

OSU-James BMT Program – Cord Blood Serious Adverse Events (SAEs)

- Both experienced sudden development of chest pain, shortness of breath, respiratory failure and pulmonary edema associated with severe heart failure (EF to 10-25%, elevated troponin), along with hematuria and acute renal failure requiring dialysis
 - Expedited reporting to CTN and NMDP
 - 1st patient felt to be chemo toxicity, patient recovered fully; 2nd raised red flags to all but unfortunately died d23 of cardiac failure
 - Both patients received 1 RBC-depleted and 1 RBC-replete CBU and both were diluted 1:1 but not washed (**NOTE** - Washing cords was actually not allowed on 05-DCB)
- May 7, 2009 – CIBMTR issues safety notice requiring notification of enrollment using RBC-replete CBUs, validation of wash method and signed acknowledgement by site PI, study coordinator, and processing lab

NMDP Investigation of SAEs

- **Spring 2009:** NMDP / CIBMTR performed a retrospective review of adverse reactions **reported to** NMDP looking for similar cases associated with cord blood infusion
 - 13 cases of caridomyopathy: 4 considered severe (including our 2)
 - 12/13 were associated with RBC-replete CBUs
- **Summer 2009:** multiple reports, memos & notifications
 - CTN to 0501/0602 and all PIs, clinical/regulatory/lab coordinators – requiring washing of RBC-replete CBUs
 - NMDP to cord IND PIs, TC Medical Directors & coordinators – explaining SAEs, investigation & **recommending** that transplants under NMDP BB-IND-7555 “comply with thaw, dilution and washing procedures set forth in Section 6.13.7 of the IND” but still allowing for provisions to eliminate washing if felt to have adverse impact.
 - NMDP to FDA
- **Summer-Fall 2009:** NMDP working group, detailed data for 6 of the cases: Cord info, Recipient info, Thaw & Infusion info, and Recipient Symptoms – **Unsure of output status**

Despite these efforts ...



Entrusted to operate the C.W. Bill Young Cell Transplantation Program,
including Be The Match Registry®

- Patient died of cardiac arrest within hours of bedside thaw of 2 RBC-replete CBUs
- As of March 2013, 8 reports to NMDP, 2 fatalities, all received at least 1 RBC-replete CBU, all (?) not washed

March 27, 2013

Celia Witten, Ph.D., M.D.
Office Director, OCTGT
Center for Biologics Evaluation & Research
Food & Drug Administration
1401 Rockville Pike (HFM 700)
Rockville, MD 20852-1448

**RE: BB-IND#: 7555-0095
IND Safety Report
NMDP Recipient Identification Number 616-970-3**

Dear Dr. Witten:

This is to inform you (in triplicate) of a fatal adverse event which occurred with a cord blood transplant recipient through the National Marrow Donor Program® (NMDP) participating in the NMDP protocol under IND#7555: "A multicenter access and distribution protocol for unlicensed cryopreserved cord blood units (CBUs) for transplantation in pediatric and adult patients with hematologic malignancies and other indications (10-CBA)". This recipient experienced a cardiac arrest following double cord blood unit infusion and was not able to be resuscitated.

FACT / JACIE Standards – 5th Edition

- **STANDARD:** *B7.4 There shall be a policy addressing safe administration of cellular therapy products.*
 - **Guidance:** “For cord blood units, the NMDP recommends the washing procedure in Appendix F of the 0501 protocol, available at www.bmtctn.net, and requires washing of red cell replete CB units due to unexpected adverse events.”
- **STANDARD:** *B7.4.1.1 Cord blood units that have **not** been red cell reduced **shall** be ~~diluted and/or washed~~.*
- **STANDARD:** *B7.4.1.2 Cord blood units that **have** been red cell reduced **should** be ~~diluted and/or washed~~.*
 - **Guidance:** “There have been documented adverse events related to the administration of cord blood units containing red blood cells. Clinical Programs need to determine the appropriate volume, DMSO (and other additives), and red cell load for recipients. These Standards require dilution and/or washing of cellular therapy products that have not been red cell reduced, and this practice is also recommended for products that have been red cell reduced.”

Better Dissemination to Key Individuals is Needed !

- Plan to post Safety Alerts on Beacon™ along with other materials from this session (pending NMDP authorization)
- Attend the Beacon™ session – QOps #9 Saturday 0730-0830

The screenshot shows the ISCT BEACON file sharing platform interface. The header includes the ISCT BEACON logo, navigation icons, and a search bar. The main content area displays a list of folders with their respective update dates and descriptions.

ISCT BEACON Suite of Navigational Tools

Search Files

All Files > BEACON > ISCT Regulatory, Quality and O...

Upload - New... - More -

File sharing platform for SOPs, validations, translational development, regulatory issues, and accreditation.

- Regulatory and Accreditation** (Updated Nov 25, 2013 by BEACON Administrator) 83
References, templates, and other tools for meeting standards and regulations around the world. Content includes: regulations, guidance documents, regulatory submission templates, gap analysis tools, and donor screening tools.
- Validation, Verification, and Qualification** (Updated Nov 25, 2013 by Joe Mierski) 3
Plans, results, references, and other tools for validations of all types. Content includes: protocols and test plans; results; facility, equipment and supply records; references to published validations; and method comparisons.
- Translational Development** (Updated Nov 5, 2013 by BEACON Administrator) 0
Templates, checklists, guidance documents, and other tools for moving from the research lab to regulatory submission. Content includes: checklists, templates, and guidance documents.
- SOPs and Forms** (Updated Nov 4, 2013 by ISCT Administrator) 9
SOPs, forms, and other "How to" tools. Content includes: policies, procedures, processes, checklists, data forms, batch records, staff records, facility records, quality plans, and audit forms.

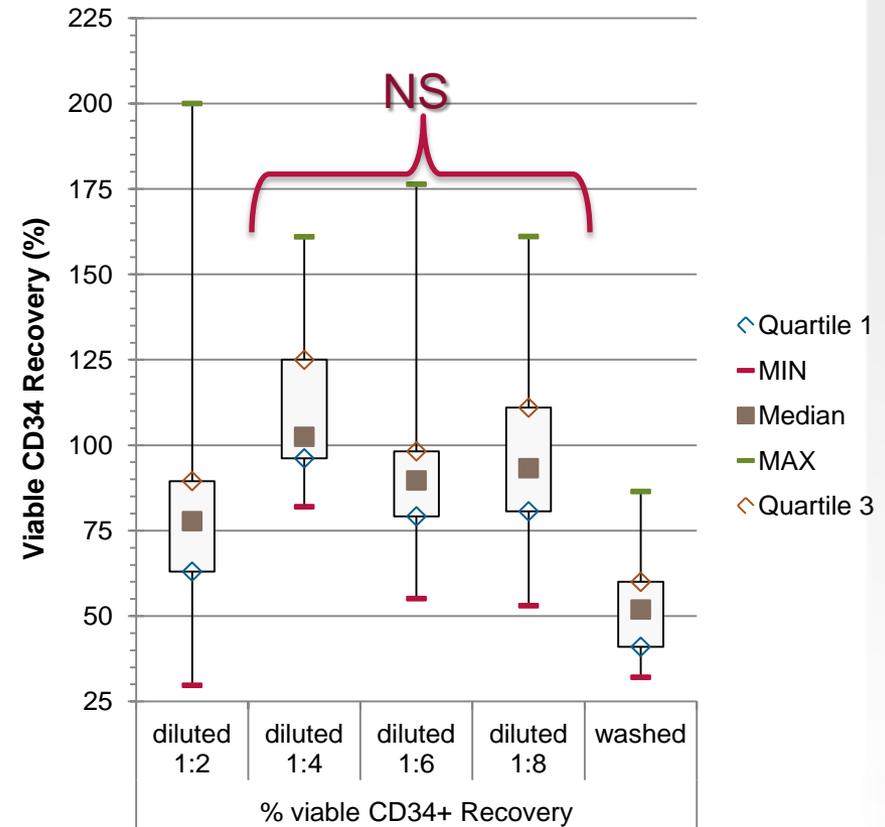
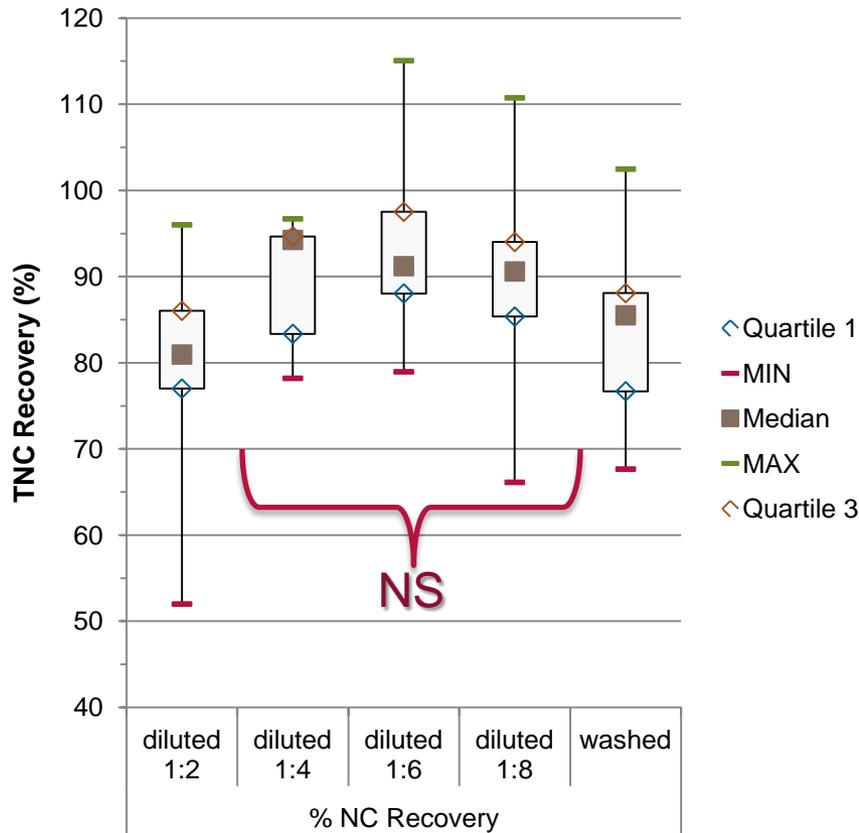
OSU Cell Therapy Laboratory (CTL): Cord Blood Processing at Thaw

- Different options for thawing cords – Goal to minimize exposure of fragile cells to DMSO to minimize cell death, to remove or dilute potential toxic molecules (RBC stoma, free hemoglobin, other)
 - Bedside thaw – no longer done at OSU (pre-2007)
 - Thaw and dilute into iso-osmotic solution (Dec 2007-current, but fold-dilution has varied over time)
 - Thaw and dilute into iso-osmotic solution, followed by centrifugation, removal of supernatant and resuspension (Dec 2008-current)
- Both cord blood banks and transplant centers need to validate thaw methods – sometimes these will conflict
- CTL sets processing targets (“criteria”) that we use in real time to inform physicians of product quality
- The true quality is in engraftment data, but this can get complicated

OSU CTL: Cord Blood Thaw Methods – Processing & Clinical Outcomes

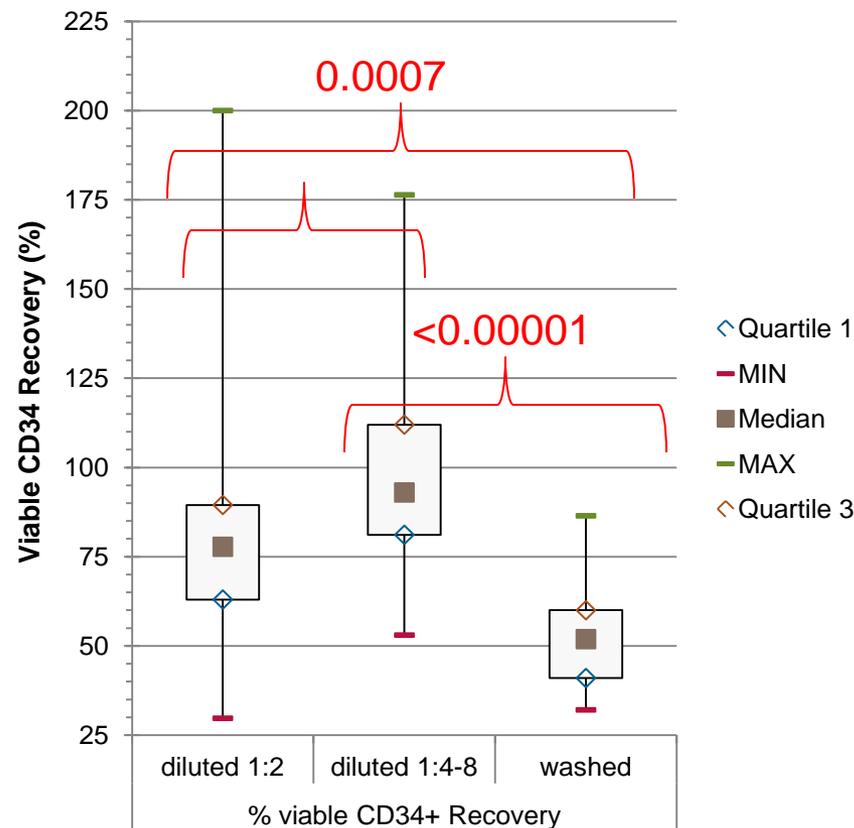
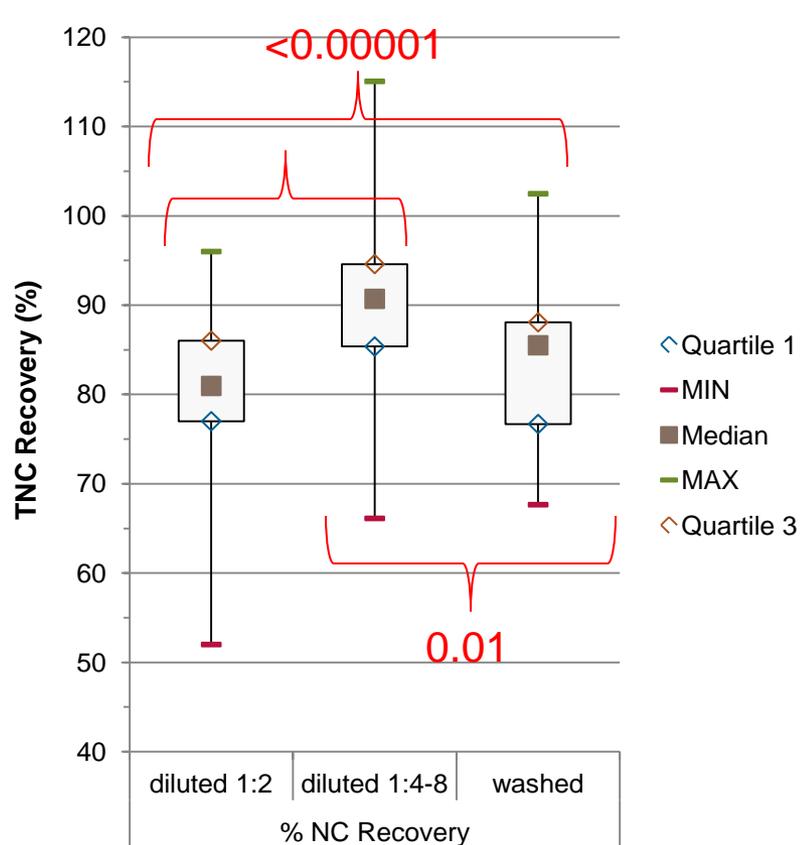
		ALL	Diluted 1:2	Diluted 1:6	Diluted 1:8	Washed
n		119	35	17	49	13
dates		Dec07- Jun12	Dec07- Feb10	Feb10- Jun12	Feb10- Jun12	Dec08- Jun12
% TNC Recovery	(mean ±SD)	87±10	80±10	94±10	90±9	83±10
	(median)	87	81	91	91	86
TNC Failure Rate (< 60%)	(%)	2%	6%	0	0	0
% vCD34 Recovery	(mean ±SD)	88±29	79±29	96±31	97±22	54±17
	(median)	84	78	90	93	52
vCD34 Failure Rate (<35%)	(%)	2%	3%	0	0	8%
% Trypan Viability	(mean ±SD)	78±6	78±8	78±5	80±5	73±5
	(median)	79	78	77	80	71
Viability Failure Rate (<60%)	(%)	0	0	0	0	0
ANC Engraftment	(mean ±SD)	25±14	21±12	29±18	25±13	28±12
	(median)	21	21	21	21	24
Delayed Engraftment Rate (>d42)	(%)	33%	21%	44%	39%	42%
PLT Engraftment	(mean ±SD)	54±49	71±82	52±33	46±24	42±26
	(median)	41	42	46	41	40
Delayed Engraftment Rate (>d60)	(%)	25%	30%	36%	18%	18%
Adverse Reaction Rate	(%)	5.9%	11.4%	0	2%	7.7%

Comparison of All Thaw Methods – Cell Recoveries (includes pediatric patients)



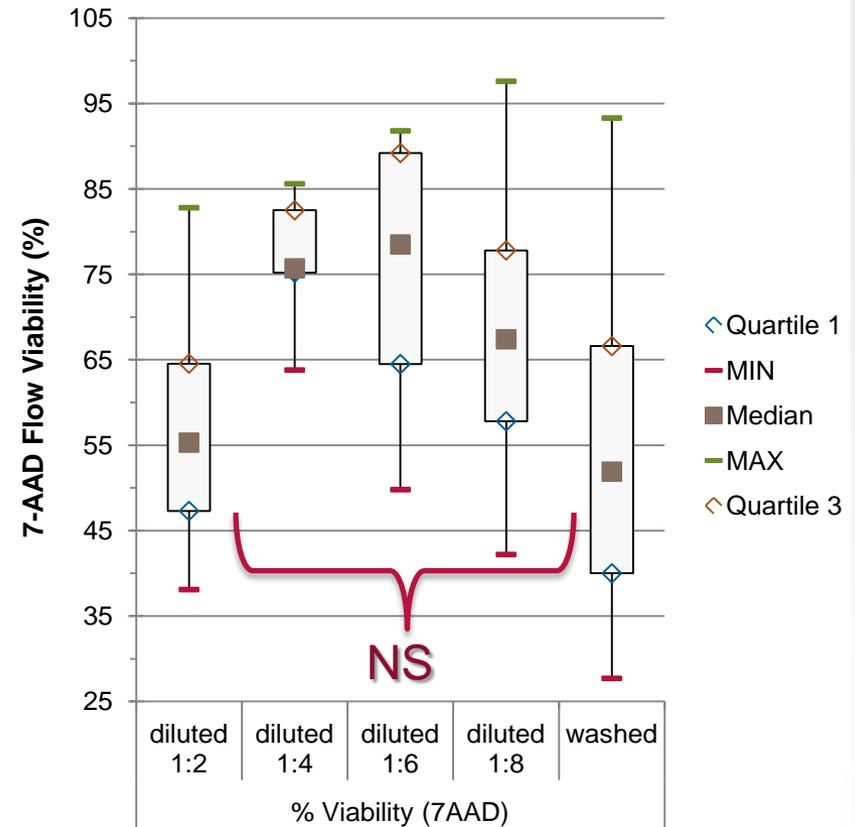
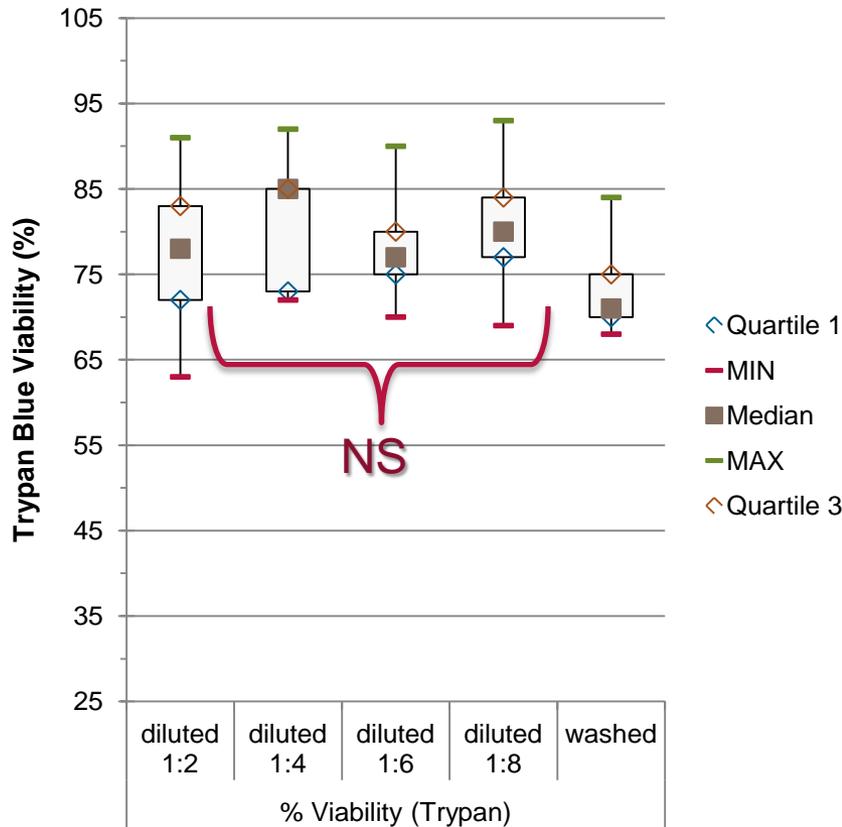
No statistically significant differences in means by t-test between 1:4, 1:6 and 1:8 dilutions (Hi-Dilute); many significant differences between these dilutions and washed or diluted 1:2 (Lo-Dilute).

Comparison of Lo-Dilute, Hi-Dilute and Wash – Cell Recoveries (includes pediatric patients)



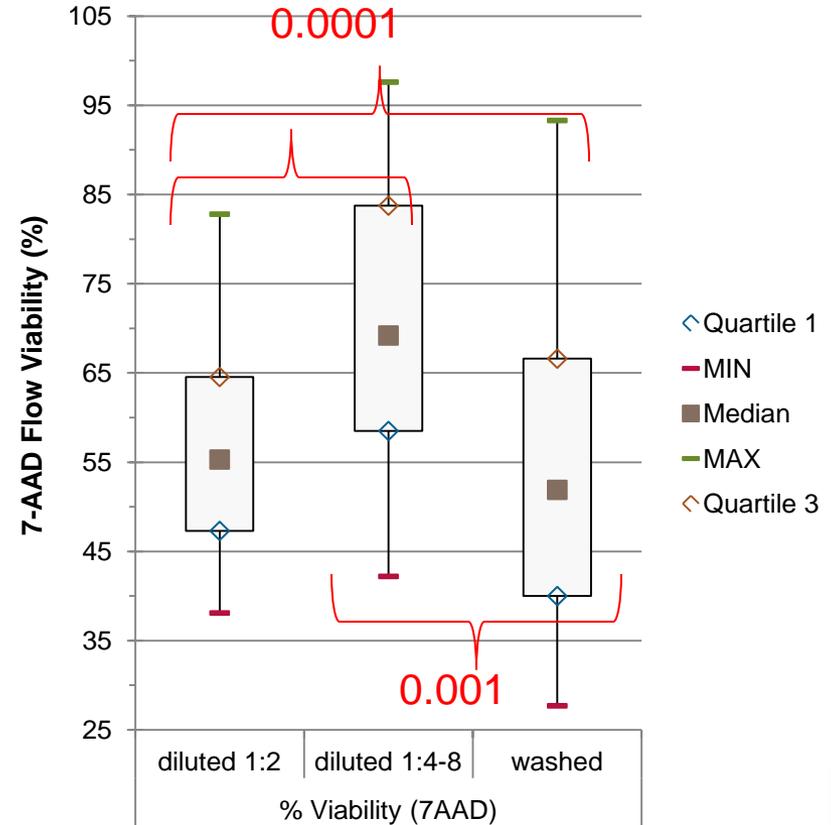
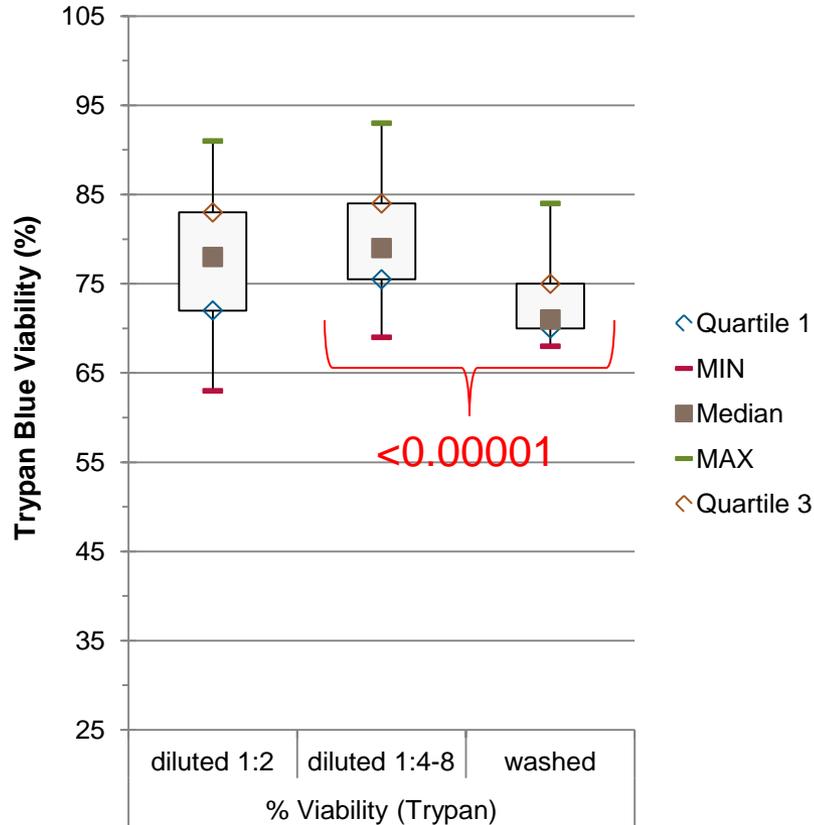
Statistically significant differences in means by t-test between all three methods. **Hi-Dilute thaw method gives best recoveries.**

Comparison of All Thaw Methods – Cell Viabilities (includes pediatric patients)



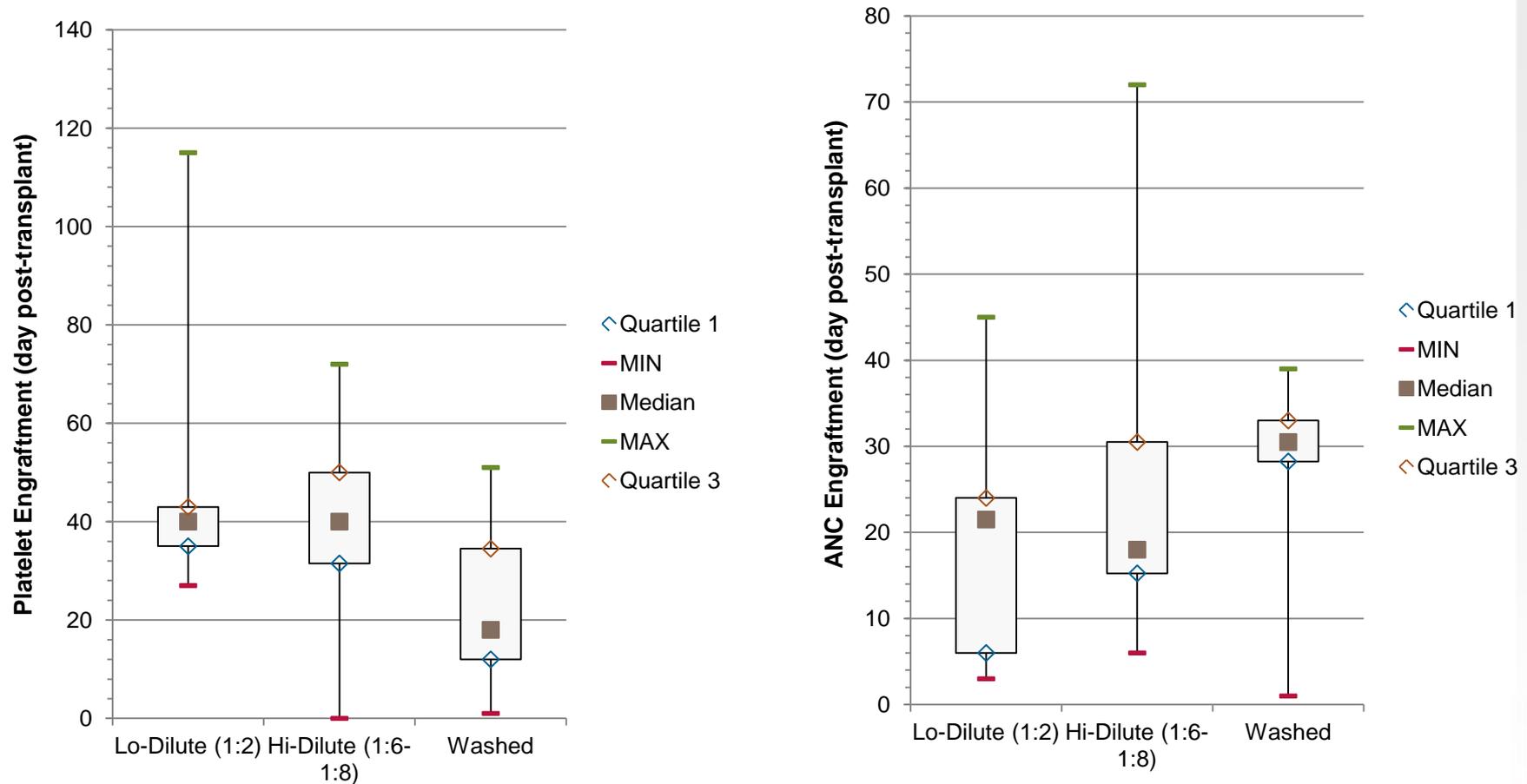
No statistically significant differences in means by t-test between 1:4, 1:6 and 1:8 dilutions (Hi-Dilute); many significant differences between these dilutions and washed or diluted 1:2 (Lo-Dilute).

Comparison of Lo-Dilute, Hi-Dilute and Wash – Cell Viabilities (includes pediatric patients)



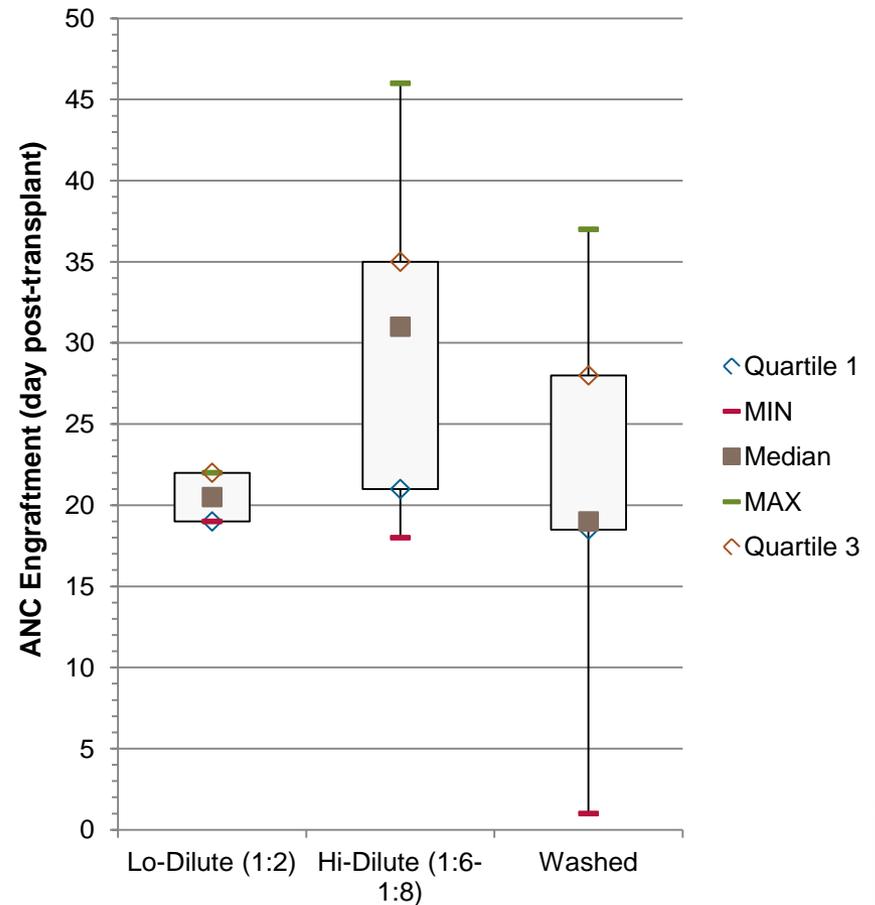
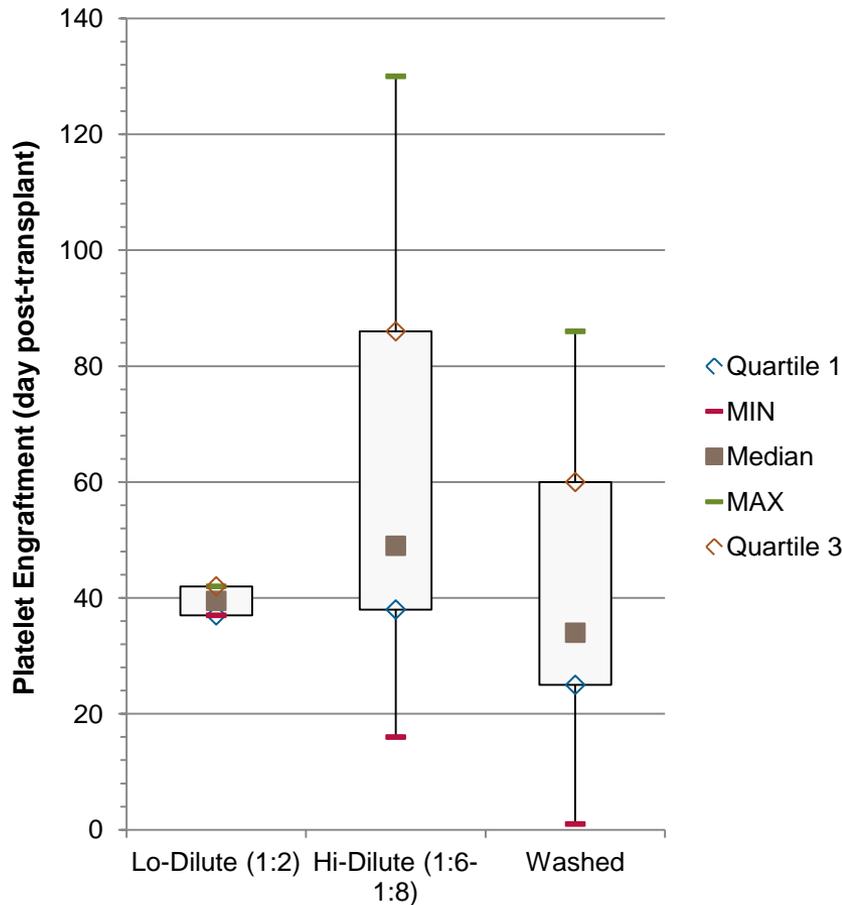
By trypan (light microscope) statistically significant difference in mean by t-test between wash and Hi-Dilute. Statistically significant differences between all three methods by flow. **Hi-Dilute thaw method gives best viabilities.**

Comparison of Lo-Dilute, Hi-Dilute and Wash – Engraftment, Non-Ablative (EXCLUDES pediatric patients)



No statistics performed; patient numbers low – 11 Lo-Dilute, 30 Hi-Dilute, only 3 Washed;

Comparison of Lo-Dilute, Hi-Dilute and Wash – Engraftment, Ablative (EXCLUDES pediatric patients)



No statistics performed; patient numbers low – 3 Lo-Dilute, 18 Hi-Dilute, only 3 Washed

Upfront Communication with Cord Blood Banks

- Was the CBU RBC-depleted?
 - NO: “plasma-reduced”, “volume reduced” or “whole blood”
→ **We must wash the CBU**
 - YES: “depleted”, “reduced”, “sedimented” or “buffy coat”
→ **We thaw/dilute the CBU – start with 1:8 dilution, but can reduce to 1:4 for larger CBUs / smaller patients or vol. overload**
 - YES BUT some “buffy coat” CBUs still contain a high level of RBCs and high volume
→ **For ALL pediatric patients or if CBU volume is >40mL, we record the RBC volume; often not available, request vol. or HCT from NMDP or bank → must wash if >2mL RBCs/kg (ped) or >20mL RBCs**
- Was CT performed on an attached segment?
 - If this cannot be confirmed, repeat CT must be performed locally
 - If attached segments – done at receipt, prior to admission
 - **If no attached segments – done at thaw – 3h TAT, 4h expiration**

Upfront Communication with Transplant Physicians

- We allow different post-thaw processing based on the cord characteristics or patient status (size, volume overload, etc.)
- CTL makes recommendations, MD must approve them or modify based on patient status

HPC, CORD BLOOD RECEIPT AND PREPARATION CHECKLIST

RECIPIENT NAME/ MRN: _____

SINGLE HPC(CB) TRANSPLANT DOUBLE HPC(CB) TRANSPLANT

RECIPIENT BODY WEIGHT: _____ kg

Completed by: Initials/ Date: _____

Reviewed by: Initials/ Date: _____

<p>HPC(CB)#1: CTL Alpha #: _____ Local CBB ID #: _____ NMDP ID #: _____ Identity verified by/ date: _____ Tech #1 _____ Tech #2 _____</p> <p>Product / Container Appearance: # bags: _____ # compartments: _____ # segments: _____ # / type ports: _____ # vials: _____ type of sample: _____ vial storage location: _____ <input type="checkbox"/> Normal <input type="checkbox"/> Abnormal: CD34* / kg _____ x 10⁵ NC/ kg _____ x 10⁷ Volume: _____ mL Storage Location: _____ Collection Date: _____ Degree of Match: _____</p> <p><u>Was product RBC-reduced prior to cryopreservation?</u> <input type="checkbox"/> NO (e.g. plasma/ volume reduced, whole blood) <input type="checkbox"/> YES (e.g. RBC-reduced/ depleted/ sedimented, buffy coat) If product is for a pediatric patient <u>or</u> if volume > 40 mL, record cryopreserved RBC volume: _____</p> <p><u>Was confirmatory HLA type performed on attached segment?</u> <input type="checkbox"/> YES - HLA typing <i>NOT</i> required <input type="checkbox"/> NO - HLA typing <i>MUST</i> be performed prior to release: Performed on: <input type="checkbox"/> attached segment <input type="checkbox"/> thawed product (3 hour TAT)</p> <p><u>Is the patient on a clinical protocol?</u> <input type="checkbox"/> NO - refer to CTL flow chart for recommended processing method <input type="checkbox"/> YES - Protocol #: _____</p> <p>Processing method specified by protocol: <input type="checkbox"/> Thaw/ Wash - specified dilution: _____ <input type="checkbox"/> Thaw/ Dilute - specified dilution: _____ <input type="checkbox"/> Not specified - refer to CTL flow chart for recommended method</p> <p>Recommended processing method Estimated final Volume <input type="checkbox"/> Thaw/ Wash _____ mL <input type="checkbox"/> Thaw/ Dilute _____ mL x _____ mL x _____ = _____ mL</p>	<p>HPC(CB)#2: CTL Alpha #: _____ Local CBB ID #: _____ NMDP ID #: _____ Identity verified by/ date: _____ Tech #1 _____ Tech #2 _____</p> <p>Product / Container Appearance: # bags: _____ # compartments: _____ # segments: _____ # / type ports: _____ # vials: _____ type of sample: _____ vial storage location: _____ <input type="checkbox"/> Normal <input type="checkbox"/> Abnormal: CD34* / kg _____ x 10⁵ NC/ kg _____ x 10⁷ Volume: _____ mL Storage Location: _____ Collection Date: _____ Degree of Match: _____</p> <p><u>Was product RBC-reduced prior to cryopreservation?</u> <input type="checkbox"/> NO (e.g. plasma/ volume reduced, whole blood) <input type="checkbox"/> YES (e.g. RBC-reduced/ depleted/ sedimented, buffy coat) If product is for a pediatric patient <u>or</u> if volume > 40 mL, record cryopreserved RBC volume: _____</p> <p><u>Was confirmatory HLA type performed on attached segment?</u> <input type="checkbox"/> YES - HLA typing <i>NOT</i> required <input type="checkbox"/> NO - HLA typing <i>MUST</i> be performed prior to release: Performed on: <input type="checkbox"/> attached segment <input type="checkbox"/> thawed product (3 hour TAT)</p> <p><u>Is the patient on a clinical protocol?</u> <input type="checkbox"/> NO - refer to CTL flow chart for recommended processing method <input type="checkbox"/> YES - Protocol #: _____</p> <p>Processing method specified by protocol: <input type="checkbox"/> Thaw/ Wash - specified dilution: _____ <input type="checkbox"/> Thaw/ Dilute - specified dilution: _____ <input type="checkbox"/> Not specified - refer to CTL flow chart for recommended method</p> <p>Recommended processing method Estimated final Volume <input type="checkbox"/> Thaw/ Wash _____ mL <input type="checkbox"/> Thaw/ Dilute _____ mL x _____ mL x _____ = _____ mL</p>
--	--

Total infusion volume: HPC(CB)#1 _____ mL + HPC(CB)#2 _____ mL = _____ mL ÷ recipient weight = _____ mL/ kg

MAXIMUM infusion volume: Recipient weight _____ kg x 20 mL/ kg = _____ mL (*total volume must be below this*)

CTL Comments: _____

<p>Order for HPC, Cord Blood Infusion:</p> <p>_____ Transplant Physician Signature Date</p>	<p>HPC(CB) #1: <input type="checkbox"/> Thaw/ Wash <input type="checkbox"/> Thaw/ Dilute - per recommendation <input type="checkbox"/> Other: _____</p>	<p>HPC(CB) #2: <input type="checkbox"/> Thaw/ Wash <input type="checkbox"/> Thaw/ Dilute - per recommendation <input type="checkbox"/> Other: _____</p>
--	---	---

Acknowledgements

- CTL staff
- BMT physicians & coordinators
- Patients

Questions? Comments?

lynn.odonnell@osumc.edu